

Antibacterial Resistance and Trend of Urinary Tract Pathogens to Commonly Used Antibiotics in Kashmir Valley

BA Tantry¹, S Rahiman²

ABSTRACT

Objective: Increase in resistance pattern of urinary tract pathogens to conventional antimicrobial agents used for urinary tract infections (UTIs) is gaining the attention of many microbiologists worldwide in respect to treatment of UTIs. The aim of the present study was to obtain data on resistance patterns of pathogens responsible for UTIs to currently used antimicrobial agents in Sher-I-Kashmir Institute of Medical Sciences (tertiary healthcare hospital).

Method: A total of 2842 samples were collected from both outpatient and inpatient departments. The majority of samples in this study were midstream urine specimens, others included catheterized urine samples. Standard parameters were followed for isolation and identification of clinical isolates and further antimicrobial susceptibility test was done by Kirby Bauer disk diffusion method.

Results: Out of 2842 samples, 1980 (67%) were culture positive. *Escherichia coli* (E coli) was the most prevalent isolate (OP 63%, IP 45.5%) followed by *Klebsiella pneumoniae* (K pneumoniae) as the second commonest UTI-causing agent (OP 15.9%, IP 21.7%). High percentage of isolates showed resistance to sulfa drugs such as cotrimoxazole. First generation cephalosporins were ineffective, while aminoglycosides and third generation cephalosporins were effective against E coli, K pneumoniae, *Pseudomonas aeruginosa* (P aeruginosa), *Enterococcus faecalis* and *Staphylococcus aureus* (Staph aureus). Furthermore, this study noticed that glycopeptide drugs such as vancomycin are highly effective against E faecalis and Staph aureus UTIs.

Conclusion: This study reveals the increased trend in resistance pattern of uropathogens in the valley region. These data may aid health professionals in choosing the appropriate treatment for patients with UTI in the region and hopefully will prevent the misuse of antibiotics.

Keywords: Antibiotic sensitivity, drug resistance, *E coli*, urinary tract infection (UTI)

Resistencia Antibacteriana y Tendencia de los Patógenos de las Vías Urinarias Frente a los Antibióticos Comúnmente Usados en el Valle de Cachemira

BA Tantry¹, S Rahiman²

RESUMEN

Objetivo: El aumento del patrón de resistencia de los patógenos de las vías urinarias frente a los agentes antimicrobianos convencionales usados para las infecciones de las vías urinarias (IVU) está ganando la atención de muchos microbiólogos a nivel mundial, en relación con el tratamiento de las IVU. El objetivo del presente estudio fue obtener datos sobre los patrones de resistencia de los patógenos responsables de las IVU en el Instituto de Ciencias Médicas Sher-I-Cachemira (hospital de atención terciaria a la salud) frente a los agentes antimicrobianos de uso común.

Método: Se recogieron un total de 2842 muestras provenientes de los departamentos de pacientes externos e internos. La mayoría de las muestras en este estudio fueron especímenes de orina de mitad de micción; otros incluyeron muestras de orina cateterizada. Se siguieron los parámetros estándar para el aislamiento y la identificación de los aislados clínicos, y posteriormente se realizó la prueba de susceptibilidad antimicrobiana mediante el método Kirby-Bauer de difusión por disco.

From: ¹Department of Microbiology, College of Medicine, Al Jouf University, Saudi Arabia and ²Department of Biochemistry, College of Medicine, Al Jouf University, Saudi Arabia.

Correspondence: Mr S Rahiman, Department of Biochemistry, College of Medicine, Al Jouf University, Sakaka, Al Jouf, Saudi Arabia, PO Box 2014, Postal code: 75471. E-mail: rahimhi@gmail.com

Resultados: De 2842 muestras, 1980 (67%) fueron cultivos positivos. *Escherichia coli* (*E coli*) fue el aislado más frecuente (OP 63%, IP 45.5%) seguido por *Klebsiella pneumonia* (*K pneumonia*) como el segundo agente más común causante de IVU (OP 15.9%, IP 21.7%). Un alto porcentaje de aislados mostró resistencia a los medicamentos de sulfa, por ejemplo el cotrimoxazole. Las cefalosporinas de primera fueron ineficaces, mientras que los aminoglucósidos y las cefalosporinas de tercera generación fueron efectivas frente a *E coli*, *K pneumoniae*, *Pseudomonas aeruginosa* (*P aeruginosa*), *Enterococcus faecalis* y *Staphyococcus aureus* (*Staph aureus*). Además, en este estudio se observó que los medicamentos glicopéptidos, tales como la vancomicina, son altamente efectivos frente a las IVU por *E faecalis* y *Staph aureus*.

Conclusión: Este estudio revela un aumento en la tendencia del patrón de resistencia de los uropatógenos en la región del Valle. Estos datos pueden ayudar a los profesionales de la salud a escoger el tratamiento apropiado para los pacientes con IVU en la región, y es de esperar que asimismo ayuden a prevenir el uso inadecuado de antibióticos.

Palabras claves: Sensibilidad antibiótica, resistencia a los medicamentos, *E coli*, infección de las vías urinarias (IVU)

West Indian Med J 2012; 61 (7): 704

INTRODUCTION

The most common human bacterial infections in the community as well as in hospital settings are urinary tract infections [UTIs] (1–2). Every year about 150 million people worldwide are affected by UTIs at a cost of about US\$6 billion (3). Urinary tract infections may involve only the lower urinary tract or may involve both the upper and lower tract. Previous studies indicated that more than 80% of uncomplicated UTIs are solely due to *Escherichia coli* [*E coli*] (4). The manifestations of UTIs are due to an interplay of the predisposing host factors and the virulence factors released by the pathogens. Congenital anomalies of the urinary tract, urinary tract obstruction, pregnancy, catheterization, instrumentation and diabetes mellitus are considered some of the predisposing factors for UTI (5), whereas virulence factors include neutrophil activation, haemolysin, adhesions and capsular polysaccharide (6). Frequently, broad-spectrum antibiotics are used to treat UTIs, however, narrow spectrum antibiotics may be suitable because of resistance concerns (7, 8). Fluroquinolones are preferred as initial antimicrobial agents because of their low rate of resistance and high clinical cure rates (9, 10). The exhaustive use of antibiotics leads to development of antibiotic resistance which becomes the major problem in treatment of UTIs globally (11).

The resistance pattern of community acquired UTI pathogens has not been studied extensively (9). Over the past years, the aetiology and antibiotic resistance pattern of uropathogens in UTIs has been changing in both community and in healthcare centres (12, 13). However, in India, the data on aetiology and antibiotic resistance pattern were not enough to treat community acquired UTIs. Hence the aim of the current study is to investigate the relative role of uropathogens and their antibacterial resistance patterns among both inpatient and outpatient departments of Sher-I-Kashmir Institute of Medical Sciences (tertiary healthcare hospital), India, to antimicrobial agents currently used in the treatment of UTI.

SUBJECTS AND METHODS

The study was conducted during February 2007 to November 2007 on patients attending the outpatient and inpatient departments of Sher-I-Kashmir Institute of Medical Sciences, Soura, Srinagar, India. A total of 2842 samples were collected from both outpatients and inpatients. The patients were provided with a sterile and dry wide mouth container and were asked to collect 10–20 ml of urine. The majority of samples were first early morning midstream urine specimens; a few inpatient samples were from urinary catheters.

Blood agar and MacConkey agar plates were inoculated with semiquantitative urine culture using a calibrated loop (14). Distinguishing between the genuine infection from contamination was done as per Kass recommendations (15). Culture of a single bacterial species from the urine sample at a concentration of > 10⁵ cfu/ml was considered as significant monomicrobial bacteriuria. Out of a total of 2842 samples, 1980 samples (750 males and 1230 females) were identified as culture proven urine isolates. Only a single positive culture per patient was included in the analysis. Standard biochemical procedures were followed to identify the significant pathogens (16).

Antibiotic susceptibility test of clinical isolates was done by the Kirby Bauer disk diffusion method (17). Antimicrobial agents tested were gentamycin 10 mcgm per disk, cephalexin 30 mcgm, cefixime 30 mcgm, nitrofurantoin 30 mcgm, amikacin 30 mcgm, ciprofloxacin 5 mcgm, vancomycin 30 mcgm (only for *E faecalis* and *Staph aureus*) and cotrimoxazole 1.25 mcgm (Hi Media, India).

SPSS (Windows Version 17.0) software was used for descriptive analysis.

RESULTS

A total of 2842 urine samples were analysed as per standard bacterial isolation and identification methods, of which 1980 (67%) samples were culture positive; 1100 (56%) were from

inpatients (IPs) [530 males and 570 females] and 880 (44%) samples were from outpatients (OPs) [220 males and 660 females].

The overall species distribution is shown in the Figure. The current study reveals that the most frequently isolated species were *E coli* (OP 63%, IP 45.5%) followed by *K pneumoniae* (OP 15.9%, IP 21.7%). In contrast, *P mirabilis* showed less frequency in prevalence (OP 1.2%, IP 2.7%).

A total of eight antibiotics were tested against all isolated uropathogens in the study, among which amikacin, cefixime, ciprofloxacin and nitrofurantoin were found to be the most effective. The antibiotic resistance pattern test of isolates to a routinely used antibiotic panel to treat UTI infections is shown in the Table. Results showed that *E coli* was

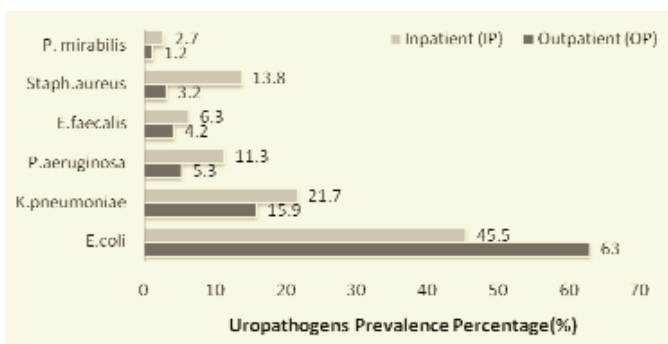


Figure: Prevalence of uropathogens among outpatients and inpatients.

Table: Antibiotic resistance pattern (%) of clinically isolated uropathogens

Microorganism	n	Antimicrobial agent	% Resistance (OP)	% Resistance (IP)
<i>E coli</i>	1054	Gentamycin	36	42
		Cephalexin	72	81
		Cefixime	16	19
		Nitrofurantoin	46	53
		Amikacin	11	13
		Ciprofloxacin	34	46
		Cotrimoxazole	76	79
<i>K pneumoniae</i>	379	Gentamycin	41	53
		Cephalexin	68	72
		Cefixime	21	25
		Nitrofurantoin	39	41
		Amikacin	17	21
		Ciprofloxacin	41	37
		Cotrimoxazole	71	78
<i>P aeruginosa</i>	171	Gentamycin	37	45
		Cephalexin	72	79
		Cefixime	14	19
		Nitrofurantoin	47	53
		Amikacin	23	27
		Ciprofloxacin	51	59
		Cotrimoxazole	73	81
<i>E faecalis</i>	106	Gentamycin	47	52
		Cephalexin	77	78
		Cefixime	23	31
		Nitrofurantoin	61	74
		Amikacin	21	33
		Ciprofloxacin	37	45
		Vancomycin	00	00
<i>P mirabilis</i>	90	Cotrimoxazole	82	87
		Gentamycin	29	30
		Cephalexin	79	81
		Cefixime	13	19
		Nitrofurantoin	33	36
		Amikacin	29	37
		Ciprofloxacin	42	51
<i>Staph aureus</i>	180	Cotrimoxazole	78	82
		Gentamycin	49	57
		Cephalexin	87	92
		Cefixime	11	17
		Nitrofurantoin	62	69
		Amikacin	33	38
		Ciprofloxacin	31	38
Vancomycin	00	00		
Cotrimoxazole	76	82		

the predominant cause of UTI, and had a higher percentage of resistance to cotrimoxazole (OP 76%, IP 79%) followed by cephalexin (OP 72%, IP 81%) and the lowest resistance to amikacin (OP 11%, IP 13%) and cefixime (OP 16%, IP 19%).

Klebsiella spp was the next most prevalent aetiological aspect of UTI in this study and displayed a slightly different resistance pattern to *E. coli*. *E. faecalis* and *Staph aureus* isolates showed the highest antibiotic susceptibility rate (100%) to vancomycin, followed by cefixime (Table). In this study, *P. mirabilis* was responsible for only about 3.9% of UTI cases (both OP and IP) and exhibited highest resistance to cotrimoxazole, cephalexin and lower resistance to cefixime (OP 13%, IP 19%).

DISCUSSION

Urinary tract infections are one of the most common infectious diseases worldwide (18–20). Due to lack of reliable indicators of UTI, early diagnosis and appropriate treatment with antibiotics are considered the most effective key factors to eliminate the uropathogens and to prevent further complications such as urosepsis and renal scarring. Urinary tract infection can be either asymptomatic or symptomatic (21). Bacteriuria with classical symptoms such as frequent urination, burning micturition and fever is referred as symptomatic UTI (22), whereas bacteriuria without classical symptoms is called asymptomatic UTI (23).

The present study shows that *E. coli* is the predominant cause of UTI amongst the outpatients as well as inpatients, *K. pneumoniae* being the next UTI-causing agent followed by *Staph aureus*, *P. aeruginosa*, *E. faecalis* and *P. mirabilis*. The frequency of *E. coli* in urine samples varies in different studies from 32% (9) to 86% (24), with intermediate values in other cases: 40% (25), 65% (26) and 68% (24). Our results (OP 63% and IP 45.5%) fit with these.

Our study indicates that *E. coli* is still the most predominant cause of UTI among in and outpatients. This corresponds with the data obtained by other investigators (1, 27, 28). Some have shown, however, that the percentage of *E. coli* is slowly declining, being replaced by other members of the Enterobacteriaceae and Enterococci (29).

The resistance pattern of *E. coli* data are similar to those obtained in other countries indicating that *E. coli* is still resistant to many antimicrobial agents (30, 31). Other species of the Enterobacteriaceae were more resistant when isolated from the hospital setting (31, 32).

In the current study, among inpatients, a high percentage of isolates showed resistance to sulfa drugs such as cotrimoxazole. First generation cephalosporins were ineffective in our study, while aminoglycosides such as amikacin and third generation cephalosporins were found to be effective against *E. coli*, *K. pneumoniae*, *P. aeruginosa*, *E. faecalis* and *Staph aureus* (Table). Furthermore, this study noticed that glycopeptide drugs such as vancomycin are highly effective against *E. faecalis* and *Staph aureus* UTIs.

Despite the abundant availability of antimicrobial agents against UTIs, it still remains one of the most common human infectious diseases (33). Antibiotic drug resistance in uropathogen may be due to overuse, abuse and at times misuse, due to wrong diagnosis and empirical prescription without urine culture (34, 35). Increased globalization could contribute to the spread of drug resistance. Appropriate knowledge on uropathogens and their antibiotic susceptibility is mandatory to ensure proper treatment of UTIs (36). Multi-drug resistant pathogens spread locally as well as globally as a part of rapid globalization (9).

Initial treatment for UTIs can be started before the availability of diagnostic test results such as urine culture and antibiotic sensitivity test. This may give an idea of the trend of antibiotic drug resistance pattern among the uropathogens and also aid in the selection of accurate drug for the appropriate treatment of UTIs.

This study reveals the increased trend of resistance pattern of uropathogens in this region, which may be due to variation in geography, misuse of drugs or wrong diagnosis. These data may also aid health professionals to choose appropriate treatment for UTI patients in the region and limit the misuse of antibiotics in the valley. Continued surveillance at both local and national levels is necessary to maintain the efficacy and safety of empirical therapy for UTIs.

REFERENCES

1. Tice AD. Short course therapy of acute cystitis: a brief review of therapeutic strategies. *J Antimicrob Chemother* 1999; **43**: 85–93.
2. Clarridge JE, Johnson JR, Pezzlo MT, Cumitech 2B: Laboratory Diagnosis of Urinary Tract Infections, (A.S Weissfeld) (Ed). Washington, DC: American Society for Microbiology; 1998.
3. Gonzalez CM, Schaeffer AJ. Treatment of urinary tract infection: what's old, what's new, and what works. *World J Urol* 1999; **6**: 372–82.
4. Rao Bhau LN, Goyal D, Chaturvedi AP, Jayasheela M, Aggarwal P. Prevalence of *Escherichia coli* sero type in urinary tract infections. *Indian J Med Microbiol* 1987; **7**: 21–5.
5. Measley RE, Levison ME. Host defense mechanisms in the pathogenesis of UTI. *Med Clin North Am* 1991; **75**: 275–86.
6. Siegfried L, Kmetova M, Puzova H, Molokacova M, Filkas J. Virulence associated factors in *E. coli* strains isolated from children with UTI. *J Med Microbiol* 1994; **41**: 127–32.
7. Schaeffer AJ. The expanding role of fluoroquinolones. *Am J Med* 2002; **113 (Suppl 1A)**: 45S–54S.
8. Biswas D, Gupta P, Prasad R, Singh V, Arya M, Kumar A. Choice of antibiotic for empirical therapy of acute cystitis in a setting of high antimicrobial resistance. *Indian J Med Sci* 2006; **60**: 53–8.
9. Gupta V, Yadav A, Joshi RM. Antibiotic resistance pattern in uropathogens. *Indian J Med Microbiol* 2002; **20**: 96–8.
10. Tankhiwale SS, Jalgaonkar SV, Ahamad S, Hassani U. Evaluation of extended spectrum beta lactamase in urinary isolates. *Indian J Med Res* 2004; **120**: 553–6.
11. Kumar MS, Lakshmi V, Rajagopalan R. Related articles, occurrence of extended spectrum beta-lactamases among Enterobacteriaceae spp. isolated at a tertiary care institute. *Indian J Med Microbiol* 2006; **24**: 208–11.
12. Manges AR, Natarajan P, Solberg OD, Dietrich PS, Riley LW. The changing prevalence of drug-resistant *Escherichia coli* clonal groups in a community: evidence for community outbreaks of urinary tract infections. *Epidemiol Infect* 2006; **134**: 425–31.

13. Kahan NR, Chinitz DP, Waitman DA, Dushnitzky D, Kahan E, Shapiro M. Empiric treatment of uncomplicated urinary tract infection with fluoroquinolones in older women in Israel: another lost treatment option? *Ann Pharmacother* 2006; **40**: 2223–7.
14. Beckford-Ball J. Related articles, management of suspected bacterial urinary tract infection. *Nurs Times* 2006; **102**: 25–6.
15. Girou E, Rioux C, Brun-Buisson C, Lobel B, Infection Committee of the French Association of Urology. The postoperative bacteriuria score: a new way to predict nosocomial infection after prostate surgery. *Infect Control Hosp Epidemiol* 2006; **27**: 847–54.
16. McNulty CA, Bowen J, Clark G, Charlett A, Cartwright K, South West G et al. How should general practitioners investigate suspected urinary tract infection? Variations in laboratory-confirmed bacteriuria in South West England. *Commun Dis Public Health* 2004; **7**: 220–6.
17. Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. *Am J Clin Pathol* 1966; **45**: 493–6.
18. McLaughlin SP, Carson CC. Urinary tract infections in women. *Med Clin North Am* 2004; **88**: 417–29.
19. Llenorroz HJ. Evidence-based management of urinary tract infections across the lifespan: management. *Clin Fam Pract* 2004; **6**: 157–73.
20. Blair KA. Evidence based urinary tract infection across the life span: current updates. *J Nurse Pract* 2007; **3**: 629–32.
21. Macejko AM, Schaeffer AJ. Asymptomatic bacteriuria and symptomatic urinary tract infections during pregnancy. *Urol Clin North Am* 2007; **34**: 35–42.
22. Hooton TM, Scholes D, Hughes JP, Winter C, Roberts PL, Stapleton AE et al. A prospective study of risk factors for symptomatic urinary tract infection in young women. *N Engl J Med* 1996; **335**: 468–74.
23. Nicolle LE. Asymptomatic bacteriuria in the elderly. *Infect Dis Clin North Am* 1997; **11**: 647–62.
24. Gupta K, Scholes D, Stamm WE. Increasing prevalence of antimicrobial resistance among uropathogens causing acute uncomplicated cystitis in women. *J Am Med Assoc* 1999; **281**: 736–8.
25. Orret FA, Shurland SM. The changing patients of antimicrobial susceptibility of urinary pathogens in Trinidad. *Singapore Med J* 1998; **39**: 256–9.
26. Barrett SP, Savage MA, Rebec MP, Guyot A, Andrews N, Shrimpton SB. Antibiotic sensitivity of bacteria associated with community acquired urinary tract infection in Britain. *J Antimicrob Chemother* 1999; **44**: 359–65.
27. Stamm WE, Hooton TM. Management of urinary tract infections in adults. *New Engl J Med* 1993; **329**: 1328–34.
28. Henry D, Ellison W, Sullivan J, Mansfield DL, Magner DJ, Dorr MB et al. Treatment of community acquired acute uncomplicated urinary tract infection with sparfloxacin versus ofloxacin. The Sparfloxacin Multi-Center UUTI Study Group. *Antimicrobial Agents and Chemotherapy* 1998; **42**: 2262–6.
29. Gruneberg RN. Changes in urinary pathogens and their antibiotic sensitivities 1971–1992. *J Antimicrob Chemother* 1994; **33 (Suppl A)**: 1–8.
30. Fluit AC, Jones ME, Schmitz FJ, Acar J, Gupta R, Verhoef J. Antimicrobial resistance among urinary tract infection (UTI) isolates in Europe: results from the SENTRY Antimicrobial Surveillance Program 1997. *Antonie van Leeuwenhoek* 2000; **77**: 147–52.
31. Cunney RJ, McNally RM, McNamara EM, Al-Ansari N, Smyth EG. Susceptibility of urinary pathogens in a Dublin teaching hospital. *Irish J Med Sci* 1992; **161**: 623–5.
32. Vromen M, van der Ven AJ, Knols AM, Stobberingh EE. Antimicrobial resistance patterns in urinary tract isolates from nursing home residents. Fifteen years of data reviewed. *J Antimicrob Chemother* 1999; **44**: 113–6.
33. Sharma SC. Understanding of pathogenic mechanisms in UTIs. *Ann Natl Acad Med Sci* 1997; **33**: 31–8.
34. Tambekar DH, Khandelwal VK. Antibigram of urinary tract pathogens. 46th Annual Conference of Association of Microbiologists of India; Osmania University, Hyderabad; December 8–10, 2005.
35. Tambekar DH, Dhanorkar DV. The prevalence and antibiogram of potential bacterial pathogens in clinical specimens. 46th Annual Conference of Association of Microbiologists of India, Osmania University, Hyderabad; December 8–10, 2005.
36. Grubenberg GN. Antibiotic sensitivities of urinary pathogens 1971–1982. *J Antimicrob Chemother* 1984; **14**: 17–23.