## Alcoholic Extract of Lotus Leaves Improves Lipid Profile in Rats with HIV Protease Inhibitor-induced Dyslipidaemia

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## ABSTRACT

**Objective:** To examine the effect of the alcoholic extract of lotus leaves (AELL) on antiretroviral treatment-induced dyslipidaemia in a rat model.

Methods: Lotus leaves were extracted by 95% ethanol. Seventy male Sprague-Dawley rats were given lopinavir/ritonavir for six weeks. At weeks 0 and 6, sera were collected for measurement of total cholesterol (TC) and triglyceride (TG). Rats meeting the criteria for dyslipidaemia were assigned to four groups and received once daily for another four weeks lopinavir/ritonavir (group A), lopinavir/ritonavir plus 0.52 g/kg AELL (group B), lopinavir/ritonavir plus 0.26 g/kg AELL (group C), or lopinavir/ritonavir plus 0.13 g/kg AELL (group D), respectively. At weeks 8 and 10, blood samples were collected again for measurement of TC and TG.

**Results:** Both TC and TG increased over time in group A during the observation period (weeks 6 to 10), however, TC and TG decreased in group B, and TG declined in group C. Neither TC nor TG could be reduced to a level near baseline.

**Conclusion:** Alcoholic extract of lotus leaves may have the potential to treat dyslipidaemia related to highly active antiretroviral treatment, but may not be potent enough to reduce TC or TG concentrations to goal levels when used alone.

Keywords: Dyslipidaemia, highly active antiretroviral therapy (HAART), lipid, lotus leaves

# El Extracto Alcohólico de las Hojas de Loto Mejora el Perfil Lipídico en Ratas con Dislipidemia Inducida por Inhibidores de la Proteasa del VIH

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## RESUMEN

**Objetivo:** Examinar el efecto del extracto alcohólico de hojas de loto (EAHL) en la dislipidemia inducida por el tratamiento de antirretrovirales en un modelo de rata.

*Métodos:* De las hojas de loto se extrajo etanol al 95%. Setenta ratas machos Sprague-Dawley recibieron lopinavir/ritonavir durante seis semanas. En las semanas 0 y 6, se recolectaron sueros para medir el colesterol total (CT) y los triglicéridos (TG). Las ratas que satisfacían los criterios diagnósticos de la dislipidemia fueron asignadas a cuatro grupos y recibieron: una vez al día diariamente por cuatro semanas lopinavir/ritonavir (grupo A); lopinavir/ritonavir más 0.52 g/kg EAHL (grupo B); lopinavir/ritonavir más 0.26 g/kg EAHL (grupo C); o lopinavir/ritonavir más 0.13 g/kg EAHL (grupo D), respectivamente. En las semanas 8 y 10, se recogieron muestras de sangre otra vez para medir CT y TG.

**Resultados:** Tanto CT como TG aumentaron con el tiempo en el grupo A durante el período bajo observación (semanas 6 a 10). Sin embargo, TC y TG disminuyeron en el grupo B, y TG disminuyó en el grupo C. Ni TC ni TG pudieron ser reducidos a un nivel cerca de la línea de base.

**Conclusión:** El extracto alcohólico de hojas de loto puede tener el potencial para tratar la dislipidemia relacionada con el tratamiento antirretroviral de gran actividad, pero puede no ser lo suficientemente potente para reducir las concentraciones de TC ó TG a los niveles de los objetivos deseados, cuando se usa solo.

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### INTRODUCTION

Highly active antiretroviral therapy (HAART) or combined antiretroviral therapy (cART) is a combination of several potent drugs against human immunodeficiency virus (HIV). Despite the clinical benefits, current HAART regimens may have some serious side effects and toxicities. Metabolic disorders, also known as lipodystrophy, including dyslipidaemia, insulin resistance, hyperglycaemia, and abnormal redistribution of body fat, are common problems with AIDS patients receiving HAART (1, 2). These changes increase the risk of developing cardiovascular diseases (3, 4). HIV protease inhibitor (PI)-containing HAART is extremely effective in decreasing viral load and has a profound impact on the clinical history of HIV in patients; meanwhile, it has also been considered as a high risk factor for the development of dyslipidaemia in this population (1, 5–7).

The treatment options for dyslipidaemia in HIV-infected patients include dietary and lifestyle modification, the use of lipid-lowering medications, and switching to a different antiretroviral drug class (8). Lifestyle changes such as smoking cessation, dietary intervention and aerobic physical activity should be introduced to patients (8, 9), but lifestyle changes may have minor effects in reducing lipids to the normal range (10-12). Switching antiretroviral drugs should only be considered when there is a viable alternative antiretroviral regimen, and the new regimen is likely to induce less dyslipidaemia than the original and is likely to possess similar or enhanced antiretroviral potency compared to the original regimen (13, 14). In addition, because of the multifactorial nature of dyslipidaemia in HIV infection, abnormal lipid profile may not be resolved simply by switching drugs (8). The common lipid-lowering agents for general patients, such as fibrates and statins, will reduce serum triglycerides and cholesterol in AIDS patients receiving cART (13, 15, 16). However, potential drug interactions between lipid-lowering agents and antiretroviral agents or other drugs may occur and lead to serious clinical consequences, such as myopathy or rhabdomyolysis (13, 17, 18).

Lotus (*Nelumbo nucifera* Gaertn) is a perennial, rhizomatous, aquatic perennial plant that belongs to the family Nelumbonaceae, and is cultivated mainly in eastern Asia and India (19). Lotus serves as an ornamental plant and its seeds, young stems, and rhizomes are consumed as food. All parts of *N nucifera* have medicinal uses (20). Lotus leaves have been used in traditional medicine to clear heat, resolve summer heat and stop bleeding (21, 22). Previous studies have demonstrated that lotus leaves are effective in hyperglycaemia, dyslipidaemia, and obesity in animal models (23–26), and also in treating dyslipidaemia in human patients (27).

To our knowledge, lotus leaves have not been used in dyslipidaemia related to HAART. In consideration of the

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highly safe properties of lotus leaves and their potential effect on dyslipidaemia, this research was conducted to evaluate the effects of alcoholic extract of lotus leaves (AELL) in treating dyslipidaemia induced with protease inhibitors (lopinavir/ ritonavir) in rats.

## SUBJECTS AND METHODS

### Plant materials and preparation of plant extracts

The leaves of Nymphaea lotus were collected from Nanning, Guangxi Province of China. The plant leaves were cleaned and dried at 60 °C to constant weight and then extracted following the procedure as previously described (28). In brief, 500 g of powdered leaves of Nymphaea lotus were soaked in 95% ethanol (2000 mL) for 72 hours and were filtered. The residue was re-extracted under the same conditions. The filtrate was then concentrated under vacuum at 30 °C. After complete evaporation, the extract was weighed and preserved at 5 °C. A total of 3000 g lotus leaves was extracted and 384 g products (weighted 12.8% of the dried leaves) were gained.

#### Animals and study procedure

Seventy male Sprague-Dawley rats weighting 180-220 g were fed with a standard rodent chow containing 4% (w/w) fat and 0.04% (w/w) cholesterol, and given a dose of 250/62.5 mg/kg lopinavir/ritonavir tablet (Kaletra, Abbott Laboratories) suspended in 0.5% sodium carboxymethyl cellulose (CMC) via oral gavage once daily. The dose of lopinavir/ritonavir was determined by referring to the previous study (29). At weeks 0 and 6, rats were fasted overnight, with free access to water, and 1 mL of blood samples was collected from the orbital plexus of rats. Sera were isolated and analysed for total cholesterol (TC) and triglyceride (TG) concentrations within six hours using enzymatic kits (Nanjing Jiancheng Bioengineering Institute, China). Dyslipidaemia was defined as an increase of  $\geq$  30% in TC or TG levels compared to week 0 levels. Rats with dyslipidaemia were randomly divided into four groups using a random number table, and received once daily for another four weeks a dose of 250/62.5 mg/kg lopinavir/ritonavir (group A), 250/62.5 mg/kg lopinavir/ritonavir plus 0.52 g/kg AELL (group B), 250/62.5 mg/kg lopinavir/ritonavir plus 0.26 g/kg AELL (group C), or 250/62.5 mg/kg lopinavir/ritonavir plus 0.13 g/kg AELL (group D). The low dose of AELL (0.13 g/kg) was equivalent to human dose based on the body surface area. The dosage of lotus leaves for human adults is 10 g daily (21). To convert the dose for humans to a dose based on surface area for rats, divide 10 g by 60 kg and then multiply by 6.2 (30, 31). This calculation resulted in a rat equivalent dose of approximately 1 g of lotus leaves per kg body weight, which equates to approximately 0.13 g of AELL per kg body weight. Lopinavir/ritonavir and AELL were suspended in 0.5% CMC. Rats were weighed once a week to adjust drug

intake. At weeks 8 and 10, blood samples were collected again, and sera were isolated and immediately stored at -80 °C until analysis for TC and TG concentrations.

#### Statistical analysis

Data are expressed as mean  $\pm$  standard deviation. Differences at each time point among groups were evaluated by oneway analysis of variance (ANOVA). Repeated-measures data were analysed by repeated-measures ANOVA and post-hoc test (Tukey HSD). A two-sided *p* value  $\leq 0.05$  was considered statistically significant. Statistics were performed using SPSS version 16.0 (SPSS Inc).

#### RESULTS

After six weeks of treatment with lopinavir/ritonavir, 51 rats met the criteria for dyslipidaemia. Two rats with lowest TG and one with lowest TC were excluded, and the remaining 48 were selected and randomly assigned into four groups, with 12 rats in each group. Analysis using the one-way ANOVA indicated that TC and TG concentrations did not significantly differ among the four groups at weeks 0 and 6 (Tables 1 and 2).

Analysis of the data obtained from weeks 6 to 10 using the repeated-measures ANOVA showed that both TC and TG concentrations in group A increased over time (p < 0.001 and p < 0.01, respectively), however, TG in groups B and C and TC in group B declined over time (p < 0.001, p < 0.01, p < 0.001, respectively). Total cholesterol slightly declined in group C but did not reach the level of statistical significance (p =0.096). Both TC and TG did not obviously vary in group D (Tables 1 and 2). The test of between-subject effects of the repeated-measures ANOVA is summarized in Table 3; the result showed that both TC and TG concentrations differed among groups (F<sub>(3, 44)</sub> = 2.898, p = 0.046 and F<sub>(3, 44)</sub> = 4.192, p =0.011, respectively).

 Table 1:
 Comparison of total cholesterol concentrations among groups at each time point and among time points within each group

Group	Week 0	Week 6	Week 8	Week 10	$F^{*}$	<i>p</i> *	
А	$2.70\pm0.34$	$3.79\pm0.39$	$4.07\pm0.46$	$4.24\pm0.39$	62.412	< 0.001	
В	$2.68\pm0.39$	$3.77\pm0.49$	$3.51\pm0.45$	$3.32\pm0.50$	37.048	< 0.001	
С	$2.65\pm0.36$	$3.78\pm0.46$	$3.72\pm0.46$	$3.57\pm0.57$	2.850	0.096	
D	$2.73\pm0.27$	$3.81\pm0.41$	$3.83\pm0.47$	$3.86\pm0.45$	0.170	0.688	
F	0.124	0.017	3.052	8.243			
р	0.946	0.997	0.038	< 0.001			

\*The statistic was calculated for repeated measures data at weeks 6, 8 and 10, and did not include those at week 0.

 Table 2:
 Comparison of triglyceride concentrations among groups at each time point and among time points within each group

Group	Week 0	Week 6	Week 8	Week 10	$F^{*}$	р
А	$0.88 \pm 0.15$	$1.53 \pm 0.36$	$1.69 \pm 0.29$	$1.74 \pm 0.27$	12.960 <sup>Å</sup>	0.003
В	$0.88\pm0.10$	$1.53\pm0.16$	$1.28\pm0.18$	$1.14 \pm 0.16$	38.883	< 0.001
С	$0.91\pm0.09$	$1.54\pm0.30$	$1.45\pm0.25$	$1.34 \pm 0.25$	$8.407^{*}$	0.008
D	$0.87\pm0.10$	$1.51 \pm 0.24$	$1.50\pm0.25$	$1.52 \pm 0.21$	0.371	0.694
F	0.226	0.027	5.562	15.237		
р	0.878	0.994	0.003	< 0.001		

\*The statistic was calculated for repeated measures data from weeks 6, 8 and 10, and did not include those at week 0.

<sup>A</sup>Mauchly's test showed that the data violated the assumption of sphericity, thus a Greenhouse-Geisser correction was made to the degree of freedom for testing the F statistic for significance.

Table 3: Test of between-subject effects of the repeated-measures ANOVA

	Source	Type III sum of squares	df	Mean square	F	р	Partial eta squared
ГС							
	Intercept	2049.448	1	2049.448	3631.763	< 0.001	0.988
	Group	4.906	3	1.635	2.898	0.046	0.165
	Error	24.830	44	0.564			
G							
	Intercept	315.299	1	315.299	1888.679	< 0.001	0.997
	Group	2.099	3	0.700	4.192	0.011	0.222
	Error	7.345	44	0.167			

ANOVA - analysis of variance; TC - total cholesterol; TG - triglyceride

Group I	Group J	TC			TG		
		Mean difference (I-J)	Std error	р	Mean difference (I-J)	Std error	р
А	В	0.501	0.177	0.034	0.336	0.096	0.006
	С	0.344	0.177	0.225	0.208	0.096	0.151
	D	0.201	0.177	0.670	0.146	0.096	0.439
В	А	-0.501	0.177	0.034	-0.336	0.096	0.006
	С	-0.157	0.177	0.812	-0.128	0.096	0.551
	D	-0.300	0.177	0.339	-0.190	0.096	0.214
С	А	-0.344	0.177	0.225	-0.208	0.096	0.151
	В	0.157	0.177	0.812	0.128	0.096	0.551
	D	-0.143	0.177	0.850	-0.062	0.096	0.916
D	А	-0.201	0.177	0.670	-0.146	0.096	0.439
	В	0.300	0.177	0.339	0.190	0.096	0.214
	С	0.143	0.177	0.850	0.062	0.096	0.916

Table 4: Tukey HSD post-hoc analysis of total cholesterol (TC) and triglyceride (TG) between groups

Further analysis by Tukey HSD post-hoc test showed that group A had higher TC and TG levels than group B (p < 0.05 and p < 0.01, respectively). No significant differences were seen in other pairwise comparisons (Table 4).

#### DISCUSSION

Dyslipidaemia in HIV/AIDS patients may be caused by HIV itself, by antiretroviral drugs, or by host factors (32). Dyslipidaemia may worsen the situation of patients receiving HAART since it has been found to be associated with increased cardiovascular events (7, 33-35). It is reported that up to 81.5% of patients on HAART for at least 12 months develop dyslipidaemia (36). Specialists have suggested that treatment should be instituted for those with previous cardiovascular disease or a high risk of cardiovascular disease (13). However, the treatment of dyslipidaemia is often complicated by potential drug interactions between lipid-lowering agents and antiretroviral agents or other drugs used in the treatment of HIV infection such as antibiotics (13). Although the underlining mechanisms of drug interactions have not yet been clearly elucidated, scientists have suggested that antiretroviral agents may competitively inhibit the CYP3A4 metabolism of lipid-lowering agents; in addition, organic anion transporting polypeptide C (OATP-1B1), apolipoprotein C-III, and sterol-regulatory element-binding protein (SREBP)-1c may also be involved (37-39). Therefore, lipid-lowering agents are required to be more carefully administered to patients on HAART compared with the general population (36). Although a few lipid-lowering drugs such as gemfibrozil, atorvastatin or rosuvastatin are relatively safe and potent for treatment of HIV-infected patients with dyslipidaemia (40, 41), they are relatively expensive for most patients in developing countries, especially those from rural areas. All of these suggest that further efforts are needed to develop effective, safe and inexpensive alternatives for HIV/ AIDS patients with dyslipidaemia.

In the present study, we determined the hypolipidaemic activity of AELL in a rat model of HIV PI-induced dyslipidaemia. The results showed that high-dose AELL could decrease serum TC and TG concentrations in rats with PI-induced dyslipidaemia. Middle-dose AELL could significantly decrease TG but only cause a slight decrease in TC, implicating that AELL may have a greater effect on TG than on TC. Although low-dose AELL could not decrease TC and TG, it was obvious that TC and TG levels in rats on low-dose AELL have remained roughly stable. On the other hand, neither TC nor TG concentration could be reduced to the level near to its baseline, even at a high dose of AELL, suggesting that AELL used alone may not be potent enough to reduce TC or TG concentrations to the goal levels, and that the active ingredients of AELL may need to be further purified and concentrated to a more appropriate concentration with stronger effects.

Combined gas/liquid chromatography-mass spectroscopy has shown that lotus leaves are rich in alkaloids and flavonoids (20, 42), both of which have been found to be associated with lipid-lowering and anti-obesity effects (43, 44). Furthermore, several isolates belonging to benzylisoquinoline alkaloids or flavonoid glycoside, gained from further partition of total alkaloids and flavonoids, were found to display significant anti-HIV activity (22). These findings suggest that the extracts of lotus leaves may have the potential to be used in HIV/AIDS patients with dyslipidaemia for two purposes, lowering lipids and inhibiting HIV.

In summary, we found that the alcoholic extract of lotus leaves decreases TC and TG concentrations in rats with dyslipidaemia induced by lopinavir/ritonavir, suggesting that it may have the potential to treat dyslipidaemia related to HAART. However, it may not be potent enough to reduce TC or TG concentrations to the goal levels when used alone.

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#### REFERENCES

- Wohl DA, McComsey G, Tebas P, Brown TT, Glesby MJ, Reeds D et al. Current concepts in the diagnosis and management of metabolic complications of HIV infection and its therapy. Clin Infect Dis 2006; 43: 645– 53.
- Carr A, Samaras K, Burton S, Law M, Freund J, Chisholm DJ et al. A syndrome of peripheral lipodystrophy, hyperlipidaemia and insulin resistance in patients receiving HIV protease inhibitors. AIDS 1998; 12: F51–F58.
- 3. Barbaro G. HIV infection, highly active antiretroviral therapy and the cardiovascular system. Cardiovasc Res 2003; **60**: 87–95.
- 4. Fichtenbaum CJ. Coronary heart disease risk, dyslipidemia, and management in HIV-infected persons. HIV Clin Trials 2004; **5:** 416–33.
- Lainka E, Oezbek S, Falck M, Ndagijimana J, Niehues T. Marked dyslipidemia in human immunodeficiency virus-infected children on protease inhibitor-containing antiretroviral therapy. Pediatrics 2002; 110: e56.
- Flint OP, Noor MA, Hruz PW, Hylemon PB, Yarasheski K, Kotler DP et al. The role of protease inhibitors in the pathogenesis of HIV-associated lipodystrophy: cellular mechanisms and clinical implications. Toxicol Pathol 2009; 37: 65–77.
- Zha BS, Wan X, Zhang X, Zha W, Zhou J, Wabitsch M et al. HIV protease inhibitors disrupt lipid metabolism by activating endoplasmic reticulum stress and inhibiting autophagy activity in adipocytes. PLoS One 2013; 8: e59514.
- Dubé MP, Stein JH, Aberg JA, Fichtenbaum CJ, Gerber JG, Tashima KT et al. Guidelines for the evaluation and management of dyslipidemia in human immunodeficiency virus (HIV)-infected adults receiving antiretroviral therapy: recommendations of the HIV Medical Association of the Infectious Disease Society of America and the Adult AIDS Clinical Trials Group. Clin Infect Dis 2003; 37: 613–27.
- Lazzaretti RK, Kuhmmer R, Sprinz E, Polanczyk CA, Ribeiro JP. Dietary interventions prevents dyslipidemia associated with highly active antiretroviral therapy in human immunodeficiency virus type 1-infected individuals: a randomized trial. J Am Coll Cardiol 2012; 59: 979–88.
- Wohl DA, Tien HC, Busby M, Cunningham C, Macintosh B, Napravnik S et al. Randomized study of the safety and efficacy of fish oil (omega-3 fatty acid) supplementation with dietary and exercise counseling for the treatment of antiretroviral therapy-associated hypertriglyceridemia. Clin Infect Dis 2005; **41:** 1498–1504.
- Manfredi R, Calza L, Chiodo F. Polyunsaturated ethyl esters of n-3 fatty acids in HIV-infected patients with moderate hypertriglyceridemia: comparison with dietary and lifestyle changes, and fibrate therapy. J Acquir Immune Defic Syndr 2004; 36: 878–80.
- Turcinov D, Stanley C, Canchola JA, Rutherford GW, Novotny TE, Begovac J. Dyslipidemia and adherence to the Mediterranean diet in Croatian HIV-infected patients during the first year of highly active antiretroviral therapy. Coll Antropol 2009; 33: 423–30.
- Feeney ER, Mallon PW. HIV and HAART-associated dyslipidemia. Open Cardiovasc Med J 2011; 5: 49–63.
- Domingos H, da Cunha RV, Paniago AM. Dyslipidaemia associated with the highly active antiretroviral therapy in AIDS patient: reversion after switching (stavudine to tenofovir and lopinavir/ritonavir to atazanavir/ritonavir). Braz J Infect Dis 2007; 11: 290–2.
- Penzak SR, Chuck SK. Hyperlipidemia associated with HIV protease inhibitor use: pathophysiology, prevalence, risk factors and treatment. Scand J Infect Dis 2000; 32: 111–23.
- Singhania R, Kotler DP. Lipodystrophy in HIV patients: its challenges and management approaches. HIV AIDS (Auckl) 2011; 3: 135–43.

- 17. Mah Ming JB, Gill MJ. Drug-induced rhabdomyolysis after concomitant use of clarithromycin, atorvastatin, and lopinavir/ritonavir in a patient with HIV. AIDS Patient Care STDS 2003; **17:** 207–10.
- Sax PE. Strategies for management and treatment of dyslipidemia in HIV/AIDS. AIDS Care 2006; 18: 149–57.
- Sridhar KR, Bhat R. Lotus a potential nutraceutical source. J Agric Technol 2007; 3: 143–55.
- Mukherjee PK, Mukherjee D, Maji AK, Rai S, Heinrich M. The sacred lotus (Nelumbo nucifera) – phytochemical and therapeutic profile. J Pharm Pharmacol 2009; 61: 407–22.
- Pharmacopoeia Commission of the People's Republic of China. Pharmacopoeia of the People's Republic of China (Part one). Beijing: China Medical Science Press; 2010: 258–59.
- 22. Kashiwada Y, Aoshima A, Ikeshiro Y, Chen YP, Furukawa H, Itoigawa M et al. Anti-HIV benzylisoquinoline alkaloids and flavonoids from the leaves of Nelumbo nucifera, and structure-activity correlations with related alkaloids. Bioorg Med Chem 2005; 13: 443–8.
- Kim AR, Jeong SM, Kang MJ, Jang YH, Choi HN, Kim JI. Lotus leaf alleviates hyperglycemia and dyslipidemia in animal model of diabetes mellitus. Nutr Res Pract 2013; 7: 166–71.
- 24. la Cour B, Mølgaard P, Yi Z. Traditional Chinese medicine in treatment of hyperlipidaemia. J Ethnopharmacol 1995; 46: 125–9.
- Siegner R, Heuser S, Holtzmann U, Söhle J, Schepky A, Raschke T et al. Lotus leaf extract and L-carnitine influence different processes during the adipocyte life cycle. Nutr Metab (Lond) 2010; 7: 66.
- Du H, You JS, Zhao X, Park JY, Kim SH, Chang KJ. Antiobesity and hypolipidemic effects of lotus leaf hot water extract with taurine supplementation in rats fed a high fat diet. J Biomed Sci 2010; 17 (Suppl 1): S42.
- Guang ZS, Wu J, Yu ZL, Liu QS, Zhang SB, Wang JJ et al. Effects of lotus leaf capsule on human's blood lipids disorder. Chin J Cardiovasc Rehab Med 2003; 12: 294–7.
- Akinjogunla OJ, Yah CS, Eghafona NO, Ogbemudia FO. Antibacterial activity of leave extracts of Nymphaea lotus (Nymphaeaceae) on methicillin resistant Staphylococcus aureus (MRSA) and vancomycin resistant Staphylococcus aureus (VRSA) isolated from clinical samples. Ann Biol Res 2010; 1: 174–84.
- 29 Waring JF, Ciurlionis R, Marsh K, Klein LL, Degoey DA, Randolph JT et al. Identification of proteasome gene regulation in a rat model for HIV protease inhibitor-induced hyperlipidemia. Arch Toxicol 2010; 84: 263– 70.
- 30. US FDA (CDER). Guidance for industry estimating the maximum safe starting dose in initial clinical trials for therapeutics in adult healthy volunteers [Internet]. Silver Spring (MD): US Food and Drug Administration, US Department of Health and Human Services [Updated 2005 Jul 6; cited 2013 Nov 12]. Available from: http://www.fda.gov/downloads/ drugs/guidancecomplianceregulatoryinformation/guidances/ucm078932. pdf
- Reagan-Shaw S, Nihal M, Ahmad N. Dose translation from animal to human studies revisited. FASEB J 2008; 22: 659–61.
- Estrada V, Portilla J. Dyslipidemia related to antiretroviral therapy. AIDS Rev 2011; 13: 49–56.
- DAD Study Group; Friis-Møller N, Reiss P, Sabin CA, Weber R, Monforte Ad et al. Class of antiretroviral drugs and the risk of myocardial infarction. N Engl J Med 2007; 356: 1723–35.
- Ceccato MG, Bonolo PF, Souza Neto AI, Araújo FS, Freitas MI. Antiretroviral therapy-associated dyslipidemia in patients from a reference center in Brazil. Braz J Med Biol Res 2011; 44: 1177–83.
- Domingos H, Cunha RV, Paniago AM, Souza AS, Rodrigues RL, Domingos JA. Rosuvastatin and ciprofibrate in the treatment of dyslipidemia in patients with HIV. Arq Bras Cardiol 2012; 99: 997–1007.
- 36. Omech B, Sempa J, Castelnuovo B, Opio K, Otim M, Mayanja-Kizza H et al. Prevalence of HIV-associated metabolic abnormalities among patients taking first-line antiretroviral therapy in Uganda. ISRN AIDS 2012; eCollection 2012: 960178.
- 37. van der Lee M, Sankatsing R, Schippers E, Vogel M, Fätkenheuer G, van der Ven A et al. Pharmacokinetics and pharmacodynamics of combined

use of lopinavir/ritonavir and rosuvastatin in HIV-infected patients. Antivir Ther 2007; **12:** 1127–32.

- Fauvel J, Bonnet E, Ruidavets JB, Ferrières J, Toffoletti A, Massip P et al. An interaction between apo C-III variants and protease inhibitors contributes to high triglyceride/low HDL levels in treated HIV patients. AIDS 2001; 15: 2397–406.
- Riddle TM, Kuhel DG, Woollett LA, Fichtenbaum CJ, Hui DY. HIV protease inhibitor induces fatty acid and sterol biosynthesis in liver and adipose tissues due to the accumulation of activated sterol regulatory element-binding proteins in the nucleus. J Biol Chem 2001; 276: 37514– 9.
- Henry K, Melroe H, Huebesch J, Hermundson J, Simpson J. Atorvastatin and gemfibrozil for protease inhibitor-related lipid abnormalities. Lancet 1998; 352: 1031–2.
- Singh S, Willig JH, Mugavero MJ, Crane PK, Harrington RD, Knopp RH et al. Comparative effectiveness and toxicity of statins among HIV-infected patients. Clin Infect Dis 2011; 52: 387–95.
- 42. Chen S, Wu BH, Fang JB, Liu YL, Zhang HH, Fang LC et al. Analysis of flavonoids from lotus (Nelumbo nucifera) leaves using high performance liquid chromatography/photodiode array detector tandem electrospray ionization mass spectrometry and an extraction method optimized by orthogonal design. J Chromatogr A 2012; **1227**: 145–53.
- Fan TT, Fa L, Fang F, Jiang YH. Effect of total alkaloids from lotus leaves on body mass and lipid regulation in vivo and in vitro. J Zhejiang Univ (Agric Life Sci) 2013; 39: 141–8.
- Yang JY, Hu L, Xu Y. Research progress of flavonoids in lotus leaves. Food Science 2007; 28: 554–8.