

Effects of Sulfated Polysaccharides on Tumour Biology

XZ Wu¹, D Chen²

ABSTRACT

Sulfated polysaccharides can act not only as anticoagulants but also as tumour inhibitors. Recent studies suggest that sulfated polysaccharides could affect tumour cells directly. Sulfated polysaccharides could inhibit the metastasis and proliferation of tumour cells by binding to growth factors and cell adhesion molecules. Moreover, sulfated polysaccharides could inhibit heparanase, which cleaves heparan sulfate chains of heparan sulfate proteoglycans and cause release of growth factors sequestered by heparan sulfate chains. Some sulfated polysaccharides can induce apoptosis and differentiation of tumour cells, but the mechanism is uncertain. In addition, sulfated polysaccharides can enhance the innate and adaptive immune response for tumour cells. Thus, the anti-tumour mechanism of sulfated polysaccharides can be explained, at least partly, through the effects on tumour biology directly.

Efectos de los Polisacáridos Sulfatados en la Biología Tumoral

XZ Wu¹, D Chen²

RESUMEN

Los polisacáridos sulfatados podrían actuar no solamente como anticoagulantes, sino también como inhibidores del tumor. Estudios recientes sugieren que los polisacáridos sulfatados podrían afectar directamente las células tumorales. Los polisacáridos tumorales podrían inhibir la metástasis y la proliferación de las células tumorales por medio de la unión con los factores de crecimiento y las moléculas de adhesión celular. Además, los polisacáridos sulfatados podrían inhibir la heparanasa, que rompe las cadenas de heparán-sulfato del proteoglicano de heparán-sulfato, dando lugar a la liberación de los factores de crecimiento secuestrados por las cadenas de heparán-sulfato. Algunos polisacáridos sulfatados podrían inducir la apoptosis y diferenciación de las células tumorales, pero el mecanismo es incierto. Además, los polisacáridos sulfatados podrían mejorar la respuesta inmunológica innata y adaptativa frente a las células tumorales. De este modo, el mecanismo antitumoral de los polisacáridos sulfatados pudiera explicarse – al menos parcialmente – a partir de los efectos sobre la biología tumoral directamente.

West Indian Med J 2006; 55 (4): 270

INTRODUCTION

Sulfated oligosaccharides, such as heparan, heparan sulfate, chondroitin 4-sulfate, chondroitin 6-sulfate and dermatan sulfate, are important ingredients of extracellular matrix (ECM). Recently, many sulfated polysaccharides have been extracted from bacteria, plants and animals. Patients with

cancer have an increased risk of venous thromboembolic complications (1). Clinical trials have suggested that low molecular weight heparin (LMWH) provides a survival advantage for cancer patients with established thrombosis (2, 3). Furthermore, some experimental studies suggested that the anti-thrombotic activity play an important role in the anti-tumour effects of sulfated polysaccharides. Sulfated polysaccharides could suppress the proliferation and metastasis of tumour cells by the inhibition of tissue factor, thrombin, thrombus formation and platelet aggregation (4).

However, other studies suggested that the anti-metastatic and anti-thrombotic activities of sulfated polysaccharides were unrelated (5, 6). Clinical trials have indicated that sul-

From: Cancer Hospital of Tianjin¹, Tianjin Medical University, Tianjin 300060, and Luzhou Medical College², Luzhou 646000, Sichuan Province, China.

Correspondence: Dr Xiong-Zhi Wu, Cancer Hospital of Tianjin, Ti-Yuan-Bei, Huan-Hu-Xi Road, He-Xi District, Tianjin, 300060, China. e-mail: ilwxz@163.com

fated polysaccharides influenced the survival in animals and patients with advanced malignancy favourably but without venous thromboembolism (7, 8). Besides thrombin, sulfated polysaccharides bind to a wide range of proteins, such as growth factors and cell adhesion molecules. As a consequence, it is more likely that the anti-cancer mechanisms of sulfated polysaccharides are not purely an anti-thrombotic effect. In the present review, we evaluated the anti-cancer effect of sulfated polysaccharides on tumour biology.

Inhibit Metastasis

In addition to providing structural integrity of ECM, heparan sulfate proteoglycans act as storage depot for a variety of heparan sulfate-binding proteins. Heparanase is an endoglycosaminidase that hydrolyze internal glycosidic linkages of heparan sulfate in cell surface and ECM, thus causing release of growth factors sequestered by heparan sulfate chains. Heparanase is over-expressed by most cancer cells and accelerate tumour growth and metastasis. The anti-metastatic effect of sulfated polysaccharides is partly attributed to their heparanase-inhibiting activity (4, 9, 10). In addition, sulfated oligosaccharides could bind vascular endothelial growth factor (VEGF) and fibroblast growth factor (FGF) but are unable to present VEGF and FGF to their high-affinity receptors. As a result, sulfated oligosaccharides inhibit VEGF- and FGF-mediated angiogenesis (12–18).

Some components of the ECM, such as collagen, laminin, fibrinogen, fibronectin and vitronectin, possess heparin-binding domains, which have affinities for heparins or heparin-like molecules. Interactions between heparin-like molecules on the cell surface and heparin-binding domains on fibronectin, vitronectin or laminin can enhance cell migration, whereas soluble heparins inhibit such interaction and consequently lead to inhibition of migration of tumour cells (2, 19).

Inhibit Proliferation

It is controversial whether sulfated polysaccharides could affect the proliferation of tumour cells (18, 20–24). Heparins inhibit the growth of human hepatoma HepG2 cells, whereas heparan sulfates had no effect on the growth of HepG2 cells. Neither heparins nor heparan sulfates had any effect on the growth of poorly differentiated and highly metastatic hepatoma cell lines, SK-Hep-1 and PLC/PRF/5 (21). In addition, heparins and LMWH inhibit the proliferation of rat hepatoma cell lines, FAO (25). Therefore, inhibition of proliferation may be mediated by the chemical properties of sulfated polysaccharides and the species of tumour cells.

Heparanase causes release of growth factors sequestered by heparan sulfate chains. Sulfated polysaccharides can inhibit heparanase and directly bind to growth factors to inhibit the growth of tumours (21, 25–27). Angiogenesis is a process that is critical for tumour growth. The anti-proliferative effect of sulfated polysaccharides is partly attributed

to anti-angiogenesis. In addition, fucoidan and sulfated heteropolysaccharide isolated from the red alga *Schizymenia dubyi* have anti-proliferative activity with a block observed in the G₁ phase (23, 28).

Induce Apoptosis

PI-88, a sulfated oligosaccharide, induces apoptosis of pancreatic islet carcinoma (18). B-1, a sulfated polysaccharide isolated from the culture filtrate of marine *Pseudomonas spp.*, induces apoptosis of human leukaemic cells (U937) (29). Internalized sulfated glycosaminoglycans interfere with transcription factor function and subsequently induce apoptosis of murine melanoma cells (30). Fucoidan-induced apoptosis in human lymphoma HS-Sultan cell lines is accompanied by the activation of caspase-3 and down-regulation of extracellular signal-regulated kinase pathway (24).

Induce Differentiation

Sulfate polysaccharide structures change during cell differentiation and sulfated polysaccharide-growth factor interactions may be affected by such changes. Human colon carcinoma cells (CaCo-2) undergo spontaneous differentiation in culture when grown on semi-permeable supports. A greater proportion of 2-O-sulfated iduronic acid units and a smaller amount of 6-O-sulfated glucosamine units exist in differentiated than in undifferentiated cells. The structural changes affect the binding of heparan sulfate to the long isoform of platelet-derived growth factor A chain but not to FGF2 (31). Significant alterations in the charge, size and sulfation pattern of heparan sulfate in PC12 pheochromocytoma cell accompany nerve growth factor-induced differentiation and neurite outgrowth (32). The sulfated heteropolysaccharide isolated from the red alga *Schizymenia dubyi can* induce the terminal maturation of non-small-cell bronchopulmonary carcinoma cells (NSCLC-N6) and arrest cells in the G₁ phase (33).

Immune Regulation

Most studies support that sulfated polysaccharides can enhance the innate immune response by promoting the tumoricidal activities of macrophages and natural killer cells (34–37). Antigen-presenting cells migrate into and out of tumour tissue to present tumour antigen to T-helper cells, as well as to produce cytokines, such as interleukin-1 beta and TNF-alpha that stimulate T-helper cells. As a result, T-helper cells promote the activity of cytotoxic T-cell, which has the strong cytotoxic effect on tumour cells. Sulfated polysaccharides can enhance the adaptive immune response by promoting such process (36, 38–40). Recent studies have implicated that sulfated polysaccharides recognize a range of cell adhesion systems. Sulfated polysaccharide could bind to CD₂, CD₃ and CD₄ in T lymphocytes and enhance the proliferative response of T lymphocytes (41–43).

CONCLUSION

Besides thrombin, sulfated polysaccharides bind to a wide range of proteins, such as growth factors and cell adhesion molecules. As a consequence, sulfated polysaccharides could affect the proliferation, differentiation, apoptosis and metastasis of tumour cells. In addition, sulfated polysaccharides could enhance the innate and adaptive immune response to tumour cells.

Thus, the anti-tumour mechanism of sulfated polysaccharides could be explained, at least partly, through the effects on tumour biology directly.

REFERENCES

- Smorenburg SM, Hutten B, Prins M. Should patients with venous thromboembolism and cancer be treated differently? *Haemostasis* 1999; **29**: 91–7.
- Green D, Hull RD, Brant R, Pineo GF. Lower mortality in cancer patients treated with low-molecular-weight versus standard heparin. *Lancet* 1992; **339**: 1476.
- Gould MK, Dembitzer AD, Doyle RL, Hastie TJ, Garber AM. Low-molecular-weight heparins compared with unfractionated heparin for treatment of acute deep venous thrombosis: A meta-analysis of randomized, controlled trials. *Ann Intern Med* 1999; **130**: 800–9.
- Smorenburg SM, Van Noorden CJ. The complex effects of heparins on cancer progression and metastasis in experimental studies. *Pharmacol Rev* 2001; **53**: 93–106.
- Bitan M, Mohsen M, Levi E, Wygoda MR, Miao HQ, Lider O et al. Structural requirements for inhibition of melanoma lung colonization by heparanase inhibiting species of heparin. *Isr J Med Sci* 1995; **31**: 106–18.
- Bertolesi GE, Lauria de Cidre L, Eijan AM. Growth inhibition in vitro of murine mammary adenocarcinoma cells by heparin and chemically modified heparins. *Tumour Biol* 1994; **15**: 275–83.
- Klerk CP, Smorenburg SM, Otten HM, Lensing AW, Prins MH, Piovella F et al. The effect of low molecular weight heparin on survival in patients with advanced malignancy. *J Clin Oncol* 2005; **23**: 2130–5.
- Williams DL, Pretus HA, McNamee RB, Jones EL, Ensley HE, Browder IW et al. Development, physicochemical characterization and preclinical efficacy evaluation of a water soluble glucan sulfate derived from *Saccharomyces cerevisiae*. *Immunopharmacol* 1991; **22**: 139–55.
- Miao HQ, Elkin M, Aingorn E, Ishai-Michaeli R, Stein CA, Vlodavsky I. Inhibition of heparanase activity and tumor metastasis by laminarin sulfate and synthetic phosphorothioate oligodeoxynucleotides. *Int J Cancer* 1999; **83**: 424–31.
- Parish CR, Freeman C, Brown KJ, Francis DJ, Cowden WB. Identification of sulfated oligosaccharide-based inhibitors of tumor growth and metastasis using novel in vitro assays for angiogenesis and heparanase activity. *Cancer Res* 1999; **59**: 3433–41.
- Murata J, Saiki I, Makabe T, Tsuta Y, Tokura S, Azuma I. Inhibition of tumor-induced angiogenesis by sulfated chitin derivatives. *Cancer Res* 1991; **51**: 22–6.
- Jayson GC, Gallagher JT. Heparin oligosaccharides: Inhibitors of the biological activity of bFGF on Caco-2 cells. *Br J Cancer* 1997; **75**: 9–16.
- Pisano C, Aulicino C, Vesci L, Casu B, Naggi A, Torri G et al. Under-sulfated, low-molecular-weight glycol-split heparin as an antiangiogenic VEGF antagonist. *Glycobiology* 2005; **15**: 1C–6C.
- Norrby K. Heparin and angiogenesis: A low-molecular-weight fraction inhibits and a high-molecular-weight fraction stimulates angiogenesis systemically. *Haemostasis* 1993; **23**: 141–9.
- Norrby K, Ostergaard P. Basic-fibroblast-growth-factor-mediated de novo angiogenesis is more effectively suppressed by low-molecular-weight than by high-molecular-weight heparin. *Int J Microcirc Clin Exp* 1996; **16**: 8–15.
- Norrby K, Ostergaard P. A 5.0-kD heparin fraction systemically suppresses VEGF165-mediated angiogenesis. *Int J Microcirc Clin Exp* 1997; **17**: 314–21.
- Lepri A, Benelli U, Bernardini N, Bianchi F, Lupetti M, Danesi R et al. Effect of low molecular weight heparan sulphate on angiogenesis in the rat cornea after chemical cauterization. *J Ocul Pharmacol* 1994; **10**: 273–80.
- Joyce JA, Freeman C, Meyer-Morse N, Parish CR, Hanahan D. A functional heparan sulfate mimetic implicates both heparanase and heparan sulfate in tumor angiogenesis and invasion in a mouse model of multistage cancer. *Oncogene* 2005; **24**: 4037–51.
- McCarthy JB, Skubitz AP, Qi Z, Yi XY, Mickelson DJ, Klein DJ et al. RGD-independent cell adhesion to the carboxy-terminal heparin-binding fragment of fibronectin involves heparin-dependent and -independent activities. *J Cell Biol* 1990; **110**: 777–87.
- Antachopoulos CT, Gagos S, Iliopoulos DC, Karayannacos PE, Tseleni-Balafouta S, Alevras P et al. Low-dose heparin treatment does not inhibit SW480 human colon cancer growth and metastasis in vivo. *In Vivo* 1996; **10**: 527–31.
- Zvibel I, Halay E, Reid LM. Heparin and hormonal regulation of mRNA synthesis and abundance of autocrine growth factors: relevance to clonal growth of tumors. *Mol Cell Biol* 1991; **11**: 108–16.
- Bertolesi GE, Lauria de Cidre L, Eijan AM. Growth inhibition in vitro of murine mammary adenocarcinoma cells by heparin and chemically modified heparins. *Tumour Biol* 1994; **15**: 275–83.
- Bourgougnon N, Roussakis C, Kornprobst JM, Lahaye M. Effects in vitro of sulfated polysaccharide from *Schizymenia dubyi* (Rhodophyta, Gigartinales) on a non-small-cell bronchopulmonary carcinoma line (NSCLC-N6). *Cancer Lett* 1994; **85**: 87–92.
- Aisa Y, Miyakawa Y, Nakazato T, Shibata H, Saito K, Ikeda Y et al. Fucoidan induces apoptosis of human HS-sultan cells accompanied by activation of caspase-3 and down-regulation of ERK pathways. *Am J Hematol* 2005; **78**: 7–14.
- Jemth P, Kreuger J, Kusche-Gullberg M, Sturiale L, Gimenez-Gallego G, Lindahl U. Biosynthetic oligosaccharide libraries for identification of protein-binding heparan sulfate motifs. Exploring the structural diversity by screening for fibroblast growth factor (FGF)1 and FGF2 binding. *J Biol Chem* 2002; **277**: 30567–73.
- Moczar M, Caux F, Bailly M, Berthier O, Dore JF. Accumulation of heparan sulfate in the culture of human melanoma cells with different metastatic ability. *Clin Exp Metastasis* 1993; **11**: 462–71.
- Desnoyers L. Structural basis and therapeutic implication of the interaction of CCN proteins with glycoconjugates. *Curr Pharm Des* 2004; **10**: 3913–28.
- Riou D, Collic-Jouault S, Pinczon du Sel D, Bosch S, Siavoshian S, Le Bert V et al. Antitumor and antiproliferative effects of a fucan extracted from *ascophyllum nodosum* against a non-small-cell bronchopulmonary carcinoma line. *Anticancer Res* 1996; **16**: 1213–8.
- Matsuda M, Yamori T, Naitoh M, Okutani K. Structural revision of sulfated polysaccharide B-1 isolated from a marine *Pseudomonas* species and its cytotoxic activity against human cancer cell lines. *Mar Biotechnol (NY)* 2003; **5**: 13–9.
- Berry D, Lynn DM, Sasisekharan R, Langer R. Poly (beta-amino ester)s promote cellular uptake of heparin and cancer cell death. *Chem Biol* 2004; **11**: 487–98.
- Salmivirta M, Safaiyan F, Prydz K, Andresen MS, Aryan M, Kolset SO. Differentiation-associated modulation of heparan sulfate structure and function in CaCo-2 colon carcinoma cells. *Glycobiology* 1998; **8**: 1029–36.
- Margolis RK, Salton SR, Margolis RU. Effects of nerve growth factor-induced differentiation on the heparan sulfate of PC12 pheochromocytoma cells and comparison with developing brain. *Arch Biochem Biophys* 1987; **257**: 107–14.
- Bourgougnon N, Roussakis C, Kornprobst JM, Lahaye M. Effects in vitro of sulfated polysaccharide from *Schizymenia dubyi* (Rhodophyta, Gigartinales) on a non-small-cell bronchopulmonary carcinoma line (NSCLC-N6). *Cancer Lett* 1994; **85**: 87–92.

34. Gorelik E. Augmentation of the antimetastatic effect of anticoagulant drugs by immunostimulation in mice. *Cancer Res* 1987; **47**: 809–15.
35. Gorelik E, Bere WW, Herberman RB. Role of NK cells in the anti-metastatic effect of anticoagulant drugs. *Int J Cancer* 1984; **33**: 87–94.
36. Yim JH, Son E, Pyo S, Lee HK. Novel sulfated polysaccharide derived from red-tide microalga *Gyrodinium impudicum* strain KG03 with immunostimulating activity in vivo. *Mar Biotechnol (NY)* 2005; **7**: 331–8.
37. Zhou G, Xin H, Sheng W, Sun Y, Li Z, Xu Z. In vivo growth-inhibition of S180 tumour by mixture of 5-Fu and low molecular lambda-carrageenan from *Chondrus ocellatus*. *Pharmacol Res* 2005; **51**: 153–7.
38. O'Sullivan GM, Boswell CM, Halliday GM. Langerhans cell migration is modulated by N-sulfated glucosamine moieties in heparin. *Exp Dermatol* 2000; **9**: 25–33.
39. Dziarski R. Synergistic enhancement of T-cell responses and interleukin-1 receptor expression by interleukin-1 and heparin or dextran sulfate. *Cell Immunol* 1992; **145**: 100–10.
40. Dziarski R. Enhancement of mixed leukocyte reaction and cytotoxic antitumor responses by heparin. *J Immunol* 1989; **143**: 356–65.
41. Miao B, Li J, Fu X, Ding J, Geng M. T-cell receptor (TCR)/CD3 is involved in sulfated polymannuroguronate (SPMG)-induced T lymphocyte activation. *Int Immunopharmacol* 2005; **5**: 1171–82.
42. Parish CR, McPhun V, Warren HS. Is a natural ligand of the T lymphocyte CD2 molecule a sulfated carbohydrate? *J Immunol* 1988; **141**: 3498–504.
43. Miao B, Geng M, Li J, Li F, Chen H, Guan H et al. Sulfated polymannuroguronate, a novel anti-acquired immune deficiency syndrome (AIDS) drug candidate, targeting CD4 in lymphocytes. *Biochem Pharmacol* 2004; **68**: 641–9.